

WM8- Bioinformatics Exercises

Analyses and interpretation of DNA variants

Hubert Hackl

Exercises

1. VEP, Oncotator (Funcotator)
2. MutSigCV
3. GPViz
4. IntoGen, cBioPortal, FannsDB (Condel, Cancer Genome Interpreter)
5. MutationAssessor, Mutation3D
6. Predict neoantigens (NetMHCpan 4.1)
7. Get correct qvalues for MutSigCV results of DLBCL patients, as for targeted sequencing (get_sig.R)
8. Get variant allele frequencies and compare with mapped reads in the IGV (get_VAF.R)
9. Compare MAF (VCF) files for variants on the same position (same type of variant, and same alternative allele) (compare_MAF.R)
10. Statistics and plot diagrams using R (maftools, MAF.R, stats.R)

Oncotator

<http://www.broadinstitute.org/ncatator/>

Get Started

Upload your input data below. Please note, only GRCh37/hg19 coordinates are supported and there is a maximum of 5,000 alterations per input file. If you need additional help, please visit our [help](#) page or feel free to [contact](#) us directly.

Upload Mutations

Durchsuchen... Keine Datei ausgewählt.

Submit

Upload TSV file of mutations in Oncotator [format](#).

Paste Mutations

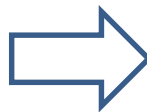


Paste data in Oncotator [format](#) or use [example](#).

Submit

jurkat_mutations.vcf

chr1	43815005	43815005	A	AT
chr1	43815008	43815008	TG	T
chr1	43815013	43815013	G	GT
chr1	43815036	43815036	G	GCC
chr1	43815041	43815041	C	G
chr1	43815047	43815047	G	GC
chr1	43815049	43815049	A	AG
chr1	115252237	115252237	T	TG



Now succeeded by Funcotator (GATK)

Oncotator

Oncotator

Toggle Annotations:

Genomic

Protein

Cancer

Non-Cancer

Specific Columns

Show 10 entries

Search:

Gene	Variant Classification	Variant Type	Hgvs Genomic Change	Hgvs Protein Change
MPL	Frame_Shift_Ins	INS	1:g.43815005_43815005insAT	ENSP00000361548;p.Leu513fs
MPL	Frame_Shift_Del	DEL	1:g.43815008delTG	ENSP00000361548;p.Trp515fs
MPL	Frame_Shift_Ins	INS	1:g.43815013_43815014dupGT	ENSP00000361548;p.Phe517fs
MPL	Intron	INS	1:g.43815036_43815038dupGCC	
MPL	Intron	SNP	1:g.43815041C>G	
MPL	Intron	INS	1:g.43815047_43815048dupGC	
MPL	Intron	INS	1:g.43815049_43815050dupAG	
NRAS	Frame_Shift_Ins	INS	1:g.115252237_115252238dupTG	ENSP00000358548;p.Ala134fs
NRAS	In_Frame_Ins	INS	1:g.115252248_115252250dupTGG	ENSP00000358548;p.His131delinsTrp
NRAS	Frame_Shift_Ins	INS	1:g.115252255_115252256dupGT	ENSP00000358548;p.Gln129fs

Showing 1 to 10 of 21 entries

Previous

1

2

3


Next

Download MAF

Download VCF

Download JSON


Variant effect predictor (VEP)


[BLAST/BLAT](#) | [VEP](#) | [Tools](#) | [BioMart](#) | [Downloads](#) | [Help & Docs](#)


VEP ▾


Web Tools


- Web Tools
 - BLAST/BLAT
 - Variant Effect Predictor**
 - Linkage Disequilibrium Calculator
 - Variant Recoder
 - File Chameleon
 - Assembly Converter
 - ID History Converter
 - VCF to PED Converter
 - Allele Frequency Calculator
 - Data Slicer
 - Variation Pattern Finder

 Configure this page

 Custom tracks

 Export data



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Variant Effect Predictor

New job

Species:

 Homo_sapiens 

Assembly: GRCh37.p13

VEP for non-human species is now only available on this site for Human (GRCh37). For other species, please visit our [main site](#).

Name for this job (optional):

Input data:

Either paste data:

Examples: [Ensembl default](#), [VCF](#), [Variant identifiers](#), [HGVS notations](#)

Or upload file:

Durchsuchen... jurkat_mutations.vcf

Or provide file URL:

Transcript database to use:

☒ Ensembl/GENCODE transcripts

☐ Ensembl/GENCODE basic transcripts

☐ RefSeq transcripts

☐ Ensembl/GENCODE and RefSeq transcripts

Additional configurations:

Identifiers ▾ Additional identifiers for genes, transcripts and variants

Variants and frequency data ▾ Co-located variants and frequency data

Variants and frequency data

Variant effect predictor (VEP)

e!GRCh37 BLAST/BLAT | VEP | Tools | BioMart | Downloads | Help & Docs Search Human... Login/Reg

VEP
Web Tools
Web Tools
BLAST/BLAT
Variant Effect Predictor
VEP analysis of jurkat_mut
Linkage Disequilibrium Calculator
Variant Recoder
File Chameleon
Assembly Converter
ID History Converter
VCF to PED Converter
Allele Frequency Calculator
Data Slicer
Variation Pattern Finder
Configure this page
Custom tracks
Export data
Share this page
Bookmark this page

Variant Effect Predictor results
Job details
Summary statistics

Category	Count
Variants processed	544
Variants filtered out	0
Novel / existing variants	424 (77.9) / 120 (22.1)
Overlapped genes	77
Overlapped transcripts	476
Overlapped regulatory features	44

Consequences (all)

- frameshift_variant: 29%
- downstream_gene_variant: 16%
- intron_variant: 15%
- non_coding_transcript_exon_variant: 7%
- upstream_gene_variant: 7%
- NMD_transcript_variant: 5%
- regulatory_region_variant: 3%
- non_coding_transcript_variant: 3%
- 3_prime_UTR_variant: 2%
- Others

Coding consequences

- frameshift_variant: 88%
- missense_variant: 6%
- synonymous_variant: 4%
- inframe_deletion: 1%
- stop_gained: 1%
- inframe_insertion: 1%
- incomplete_terminal_codon_variant: 0%
- start_lost: 0%
- protein_altering_variant: 0%
- coding_sequence_variant: 0%

Results preview
Navigation (per variant) **Filters**
Show: 1 5 10 50 All variants **Uploaded variant** is defined **Add**
Download **New job**
All: [VCF VEP TXT](#)
BioMart: [Variants](#) [Genes](#)

Show/hide columns (11 hidden)

Uploaded variant	Location	Allele	Consequence	Symbol	Gene	Feature type	Feature	Biotype	Exon	cDNA position	Scroll to see more columns	Codons	Existing variant	Feature strand	SIFT	PolyPhen	
1.43815005-43815005	T	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000372470.3	protein_coding	10/12	1582-1583	1540-1541	514	R/MX	AGG/ATGG	-	1	-	-
1.43815005-43815005	T	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000413998.2	protein_coding	10/10	1540-1541	1540-1541	514	R/MX	AGG/ATGG	-	1	-	-
1.43815008-43815009	-	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000372470.3	protein_coding	10/12	1586	1544	515	W/X	TGG/TG	COSV65243776 COSV65245195	1	-	-
1.43815008-43815009	-	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000413998.2	protein_coding	10/10	1544	1544	515	W/X	TGG/TG	COSV65243776 COSV65245195	1	-	-
1.43815013-43815013	T	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000372470.3	protein_coding	10/12	1590-1591	1548-1549	516-517	-/X	-/T	-	1	-	-
1.43815013-43815013	T	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000413998.2	protein_coding	10/10	1548-1549	1548-1549	516-517	-/X	-/T	-	1	-	-
1.43815036-43815036	CC	splice_region_variant, intron_variant	MPL	ENSG00000117400	Transcript	ENST00000372470.3	protein_coding	-	-	-	-	-	-	-	1	-	-
1.43815036-43815036	CC	frameshift_variant	MPL	ENSG00000117400	Transcript	ENST00000413998.2	protein_coding	10/10	1571-1572	1571-1572	524	R/RX	CGC/CGCCC	-	1	-	-
1.43815041-43815041	G	intron_variant	MPL	ENSG00000117400	Transcript	ENST00000372470.3	protein_coding	-	-	-	-	-	-	-	1	-	-
1.43815041-43815041	G	missense_variant	MPL	ENSG00000117400	Transcript	ENST00000413998.2	protein_coding	10/10	1576	1576	526	R/G	CGC/GGC	-	1	0.37	0
1.43815047-43815047	C	intron_variant	MPL	ENSG00000117400	Transcript	ENST00000372470.3	protein_coding	-	-	-	-	-	-	-	1	-	-

MutSigCV

https://www.broadinstitute.org/cancer/cga/mutsig_run (see key.txt)

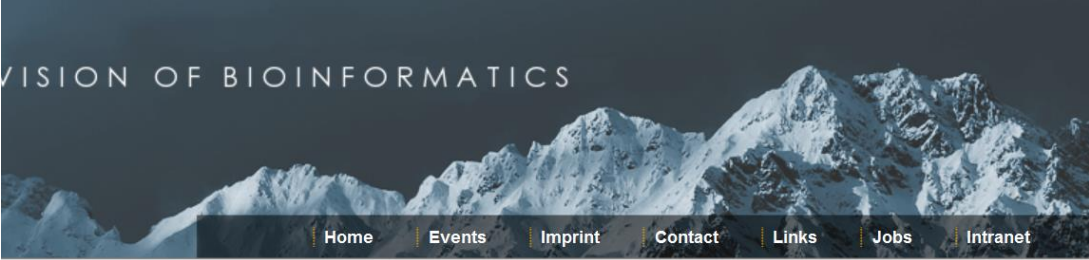
The screenshot shows the GenePattern web interface for the MutSigCV tool. The left sidebar contains navigation links for Modules, Jobs, and Files, along with search and favorite modules sections. The main panel displays the configuration form for MutSigCV, with several fields highlighted by red boxes and labeled with their respective file names:

- mutation table file***: dlcbl_prim.maf
- coverage table file***: http://genepattern.broadinstitute.org/gp/data/xchip/gpprod/shared_data/example_files/MutSigCV_1.3/exome_full192.coverage.txt
- covariates table file***: ftp://ftp.broadinstitute.org/pub/genepattern/example_files/MutSigCV_1.3/gene.covariates.txt
- output filename base***: mutation_table_dlcbl_prim
- mutation type dictionary**: http://genepattern.broadinstitute.org/gp/data/xchip/gpprod/shared_data/example_files/MutSigCV_1.3/mutation_type_dictionary_file.txt
- genome build**: hg19

At the bottom of the form, there are 'Reset' and 'Run' buttons. The footer of the page includes 'About GenePattern | Contact Us' and '©2003-2014 Broad Institute, MIT'.

GPViz

<http://icbi.at/software/gpviz/gpviz.shtml>



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GPViz:

News | Description | Documentation | License | Download | Forum

Downloads

Release	Date	Description	Instructions	Download
1.2.8	17.05.2013	GPViz ZIP package incl. test data	User Manual	download
1.2.8	17.05.2013	GPViz Webstart (1GB RAM)	User Manual	download
1.2.7	17.04.2013	GPViz ZIP package incl. test data	User Manual	download
1.2.6	27.03.2013	GPViz ZIP package incl. test data	User Manual	download

Do not update JAVA!

GeneProteinViz

File Filter View Tools Help

Load Data: Options

Load Gene Models

Load Regions

Load Variants

Load Domains

Select Gene:

ABCA7

ABL1

AKT1

ALK

APC

ARMC10

ARRB1

ATM

BRAF

C12orf11

C17orf97

CDH1

DNAH5

DPYS

EGFR

ERBB2

ERBB4

EZH2

FBXW7

FGFR2

FGFR3

Refseq_hg19.gtf

oncotorator.maf

LAML.maf

Refseq_CDD.txt

Gene ABL1 on 9.13358

5'

ABL1

Color schemes Annotations Advanced

Refseq Annotation File WM2\Mutect2-Results\Human_Refseq.txt Browse...

Ensembl Annotation File Browse...

Web links:

Gene link [http://www.ncbi.nlm.nih.gov/gene/\\${unigene_id}](http://www.ncbi.nlm.nih.gov/gene/${unigene_id})

Transcript link [http://www.ncbi.nlm.nih.gov/nuccore/\\${transcript_refseq}](http://www.ncbi.nlm.nih.gov/nuccore/${transcript_refseq})

Protein link [http://www.ncbi.nlm.nih.gov/protein/\\${protein_refseq}](http://www.ncbi.nlm.nih.gov/protein/${protein_refseq})

Genome Browser: [http://www.ncbi.nlm.nih.gov/genome/browser?db=Human&db=hg19&position=chr\\${chrom}:\\${start}-\\${stop}](http://www.ncbi.nlm.nih.gov/genome/browser?db=Human&db=hg19&position=chr${chrom}:${start}-${stop})

NM_005157

Union transcript

Union protein

NP_009297

SH3

SH2_ABL

SH2 superfamily

Plkinase_Ty

PTKc_Abl

PKc_like sup

IntOGen



FannsDB

FannsDB is a database for Functional ANnotations for Non Synonymous SNVs which contains precalculated scores for several predictors.

[Documentation](#)[Download](#)

Condel

Condel is a method to assess the outcome of non-synonymous SNVs using a CONsensus DEleteriousness score that combines various tools ([MutationAssessor](#), [FATHMM](#)). This is the second version of Condel which includes an update of the combined tools and a new web interface. If you use Condel, please cite us:

Improving the Assessment of the Outcome of Nonsynonymous SNVs with a Consensus Deleteriousness Score, Condel (2011) Abel González-Pérez and Nuria López-Bigas, American Journal of Human Genetics 10.1016/j.ajhg.2011.03.004 [Download PDF](#)

The last update of CONDEL was released on 2014. No new updates are planned for the future.

[▶ Query](#)

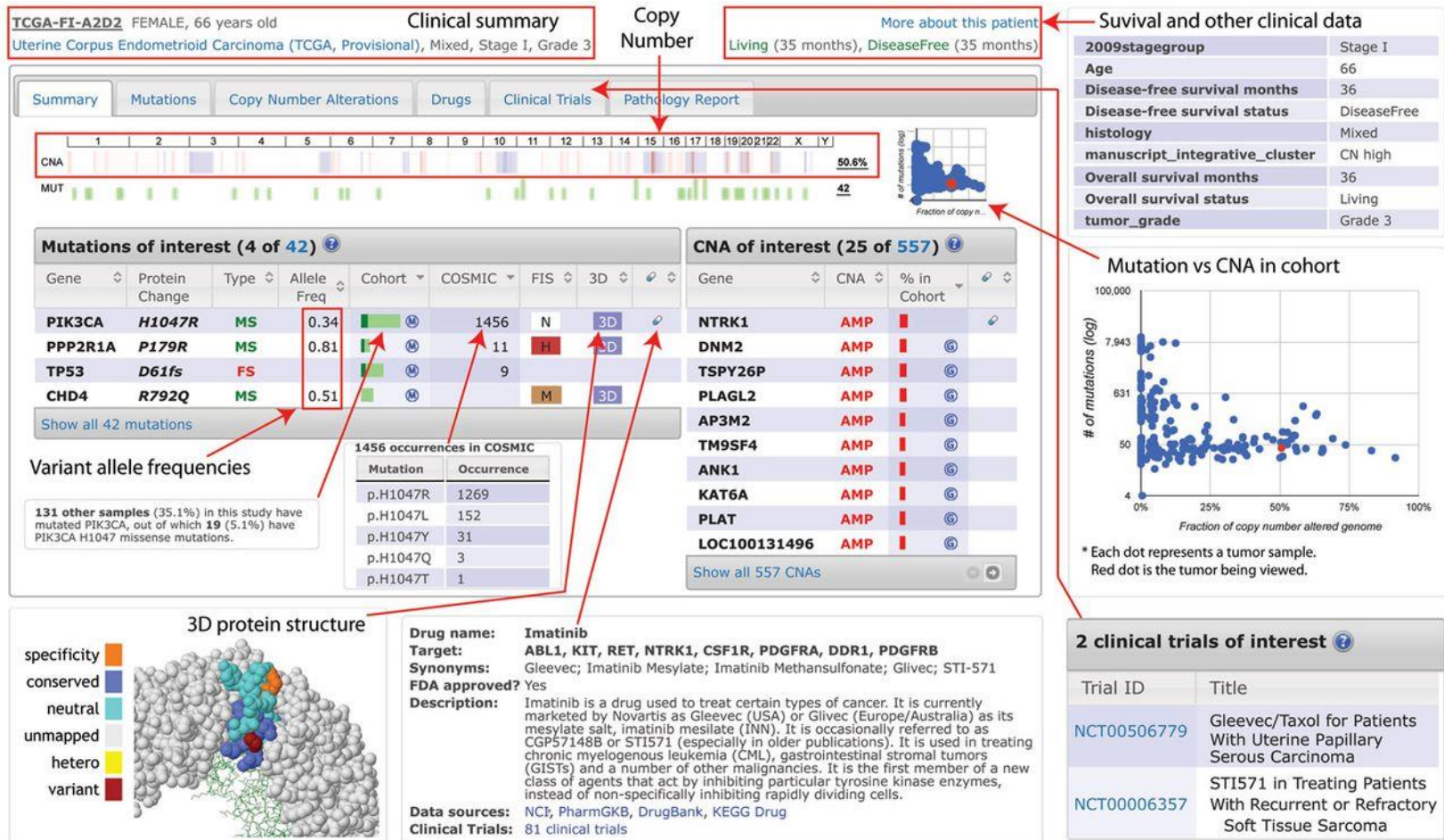
TransFIC

TransFIC (TRANSformed Functional Impact for Cancer) is a method to transform Functional Impact scores taking into account the differences in basal tolerance to germline SNVs of genes that belong to different functional classes. This transformation allows to use the scores provided by well-known tools (e.g. SIFT, Polyphen2, [MutationAssessor](#)) to rank the functional impact of cancer somatic mutations. Mutations with greater TransFIC are more likely to be cancer drivers. If you use TransFIC, please cite us:

Improving the prediction of the functional impact of cancer mutations by baseline tolerance transformation (2012) Gonzalez-Perez A, Deu-Pons J and Lopez-Bigas N. Genome Medicine. 4:89 doi:10.1186/gm390s [Read](#)

We have stopped supporting TRANSFIC. To evaluate the oncogenic potential of cancer mutations, please use the [Cancer Genome Interpreter](#).

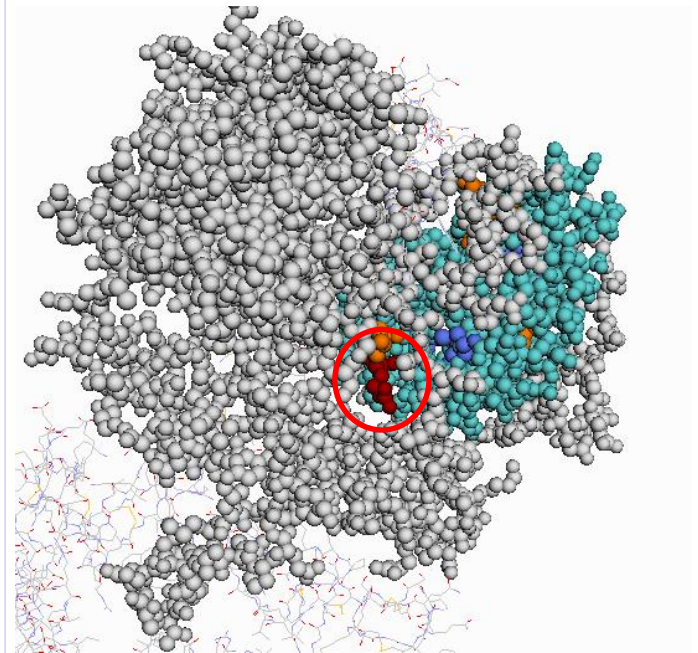
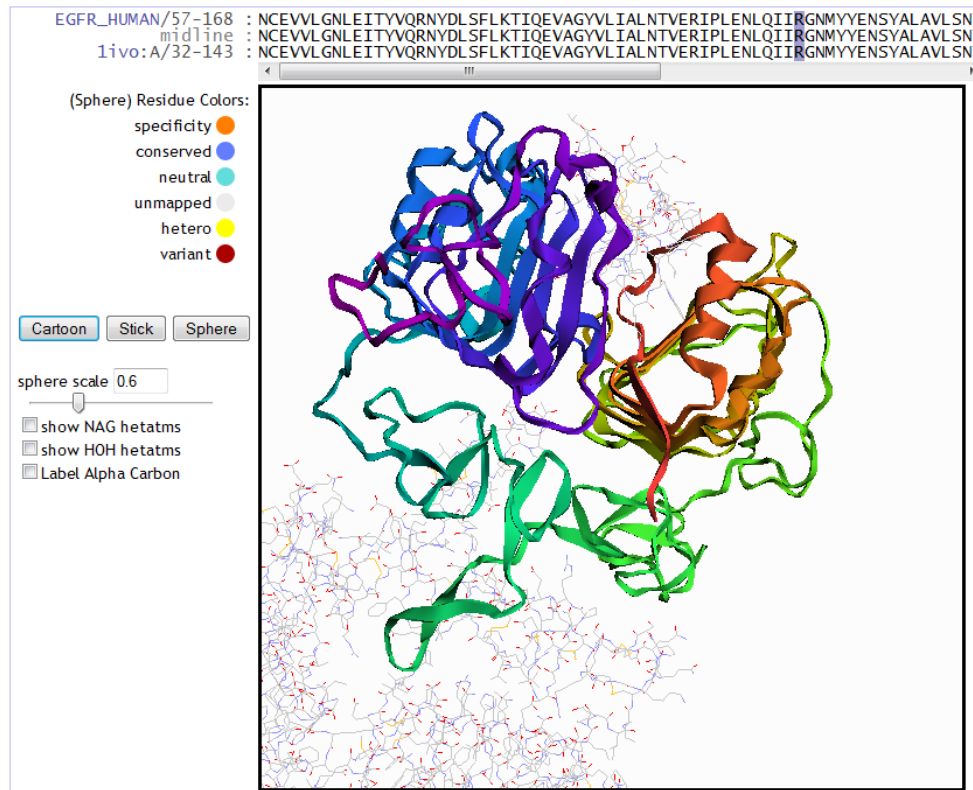
[▶ Query](#)



MutationAssessor

Chr Pos RefAlt AltAll

Mutation	AA variant	Gene	MSA	PDB	Func. Impact	FI score	Uniprot	Refseq	MSA height
2,212288939,C,G	G936A	ERBB4	msa	pdb	medium	2.4	ERBB4_HUMAN	XP_005246432	1684



mutation3D




Home Advanced About

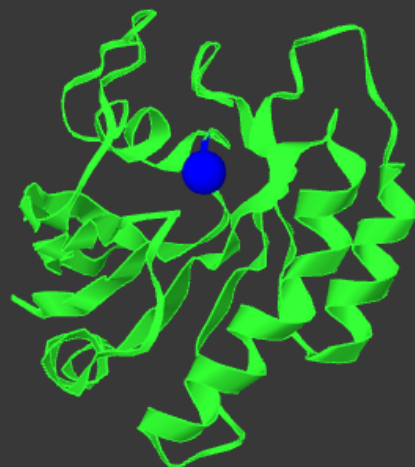
GTPase KRas

Protein Data

UniProt ID [P01116](#)
Gene KRAS
Length 189
Mutations 1
Clusters 0
Models 24

Links

 [Clustering Data File](#)
 [PyMOL Session File](#)
 [Permalink](#)



View: [Cartoon](#) [Line](#) [Ribbon](#)

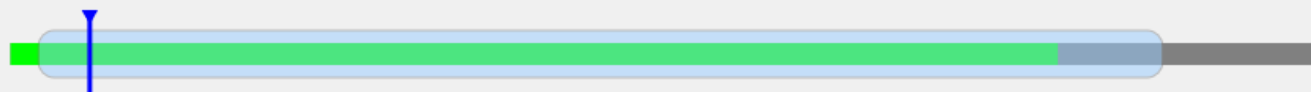
Model Information

Model: [3GFT_A_1_150](#)
Source: PDB

AA Range	1-150
Source PDB	3GFT
Coverage	79.37%
Seq. Identity	100%

Cluster Selection

No clusters were found for this model.



[Hide Domains](#)

[Select Model](#)



Cornell University
Weill Institute for Cell and Molecular Biology




Yu Group
Computational Suite

Visit the [Yu Lab website](#) for contact information and for other tools from the Yu Group Computational Suite

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NetMHCpan 4.1

Submission	Instructions	Output format	Motif viewer	Abstract	Evaluation data sets	Version history
------------	--------------	---------------	--------------	----------	----------------------	-----------------

INPUT TYPE: Fasta 

Paste a single sequence or several sequences in [FASTA](#) format into the field below:


ASTPGHTIIYEAVCLHNDRTTIP

*... or **upload** a file in [FASTA](#) format directly from your local disk:*

Durchsuchen... Keine Datei ausgewählt.

*... or **load** some sample data:*

Load Data

PEPTIDE LENGTH: 


You may select multiple lengths

8mer peptides


9mer peptides

10mer peptides

11mer peptides

SELECT SPECIES/LOCI: 

HLA supertype representative 

Select Allele(s) (max 20 per submission) 

HLA-A*01:01 (A1)

HLA-A*02:01 (A2)

HLA-A*03:01 (A3)

HLA-A*24:02 (A24)

HLA-A*26:01 (A26)

Some questions

1. How many somatic variants have you found
2. Are those point mutations, indels or other type
3. Are those synonymous, non-synonymous, or none coding
4. Changes in protein sequence? (p.V412A)
5. How many in intron, 3'UTR, cds, 5'UTR
6. Type of variant (missense mut, frame shift del, frame shift ins, etc)
7. How many variants
8. Where in the 3D structure of the protein are the variants
9. Are these known variants (dbSNP, COSMIC)
10. Is there a functional impact (SIFT, PolyPhen2)
11. Are variants cancer drivers
12. Which genes/exons are studied (info about the panel, and why this was chosen)
13. Variants per megabase of studied DNA sequence

Question

14. Which transcripts and protein isoforms are covered by the variants (NM_,NP_,ENST, ENSP)
15. Which chromosome and position of the variant
16. Reference allele, alternative allele
17. Are there common variants to previously sequenced Melanoma cell lines
18. Are some of the genes found significantly and recurrently mutated in a cohort of 28 diffuse large B cell lymphoma patients by targeted exome sequencing
19. Are there somatic variants in the Melanoma TCGA cohort patients at the same positions. What are the allele frequency in this cohort?
20. Are some variants from the same cancer signalling pathway